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Synthesis of highly oxygenated carbocyclic derivatives: decalins and cyclohexanes from sugar allyltins

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Abstract—Sugar allyltin derivatives are readily converted into the carbobicyclic compounds with precisely defined stereochemistry. Model functionalization of the carbocyclic skeleton in such precursors with the decalin structure provides either highly oxygenated bi- or monocarbocyclic derivatives.

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1. Introduction

Over the past few years, we have described a general methodology for the preparation of highly oxygenated carbobicyclic derivatives from sugar allyltins 2 and 3, which are readily available from the corresponding allylic alcohols 1 $(Fig. 1).$ $(Fig. 1).$ $(Fig. 1).$ ¹ These organometallics can be regarded as precursors of either fully oxygenated bicyclic (decalins and perhydroindanes) or monocyclic (cyclohexane and cylopentane) sugar mimetics.

The synthesis and biological properties of such monocarbocyclic derivatives are very well documented.[2](#page-5-0) However, the chemistry of the bicyclic analogues is almost unexplored. Mehta and Ramesh described a convenient route to the racemic, fully oxygenated bicyclo[4.3.0]nonanes and bicyclo[4.4.0]decanes and found that such derivatives might possess interesting inhibitory glucosidase activity.[3](#page-5-0) The highly oxygenated decalin skeleton can also be found in the structure of macrolide antibiotics such as nargenicin A, the synthesis of which in enantiomerically pure form was realized by Roush in 1996.^{[4](#page-5-0)}

Our methodology, $\frac{1}{1}$ $\frac{1}{1}$ $\frac{1}{1}$ as shown in Figure 1, allows us to obtain enantiomerically pure bicyclic derivatives with the hydrindane and decalin structures. The sugar allyltin derivatives (either primary 2 or secondary 3) undergo a controlled fragmentation to E-dienoaldehydes 5 upon treatment with a Lewis acid (the best being $ZnCl_2$),^{[5,6](#page-5-0)} while at high temperature (boiling xylene) the secondary ones 3

Figure 1. Synthesis of highly oxygenated carbobicyclic derivatives from sugar allyltins.

are converted into Z-dienoaldehyde 6 (the primary analogues 2 remain unchanged under these conditions).^{[6](#page-5-0)} These highly reactive dienes are precursors of enantiomerically

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Figure 2. Attempts to oxidize the allylic methylene group.

pure carbobicycles with well-defined structures. We were able to prepare cis-decalins 7 and trans- or cis-hydrindanes **8** in good yields ([Fig. 1](#page-0-0)).^{\dagger [,1](#page-5-0)}

Proper functionalization of the allylic system either in 7 or 8 should afford the fully oxygenated derivatives including also the analogues containing heteroatoms other than oxygen. For example, osmylation of the double bond in such carbobicyclic precursors led to the cis-diols, while epoxidation, followed by the opening of the epoxide ring provided trans-diols in good yields.⁸ However, derivatization of the allylic methylene group in such derivatives was found to be troublesome. Direct allylic oxidation (or bromination) did not give any expected product. The well-known base-induced isomerization of the correspond-ing epoxides^{[9](#page-5-0)} was also not applicable to our precursors; for example, treatment of epoxide 9 with LDA led to the rearrangement product 11 instead of the desired allylic alcohol [10](#page-5-0) (Fig. 2). 10

Herein, we report the functionalization of the decalin of type 7 leading either to highly oxygenated bicyclic or monocylic compounds.

2. Results and discussion

Adduct 12 (readily obtained from the corresponding D-gluco-configurated allyltin according to Ref. [11](#page-5-0)) was chosen as a precursor of a highly oxygenated decalin (after functionalization of the $C6-C8$ allylic system in ring A) and a highly functionalized cyclohexane. The monocyclic derivatives can be obtained from 12 either by transformation of ring A^{\dagger} or

Figure 3. Possible routes of functionalization of the bicyclic precursor 12.

B (via a Baeyer–Villiger oxidation^{[12](#page-5-0)} of the C1-carbonyl group), see Figure 3.

These two approaches which open the routes to carbocyclic sugar mimics will be discussed.

2.1. Model synthesis of highly functionalized cyclohexane derivative from adduct 12

The Baeyer–Villiger oxidation, 13 which directly converts ketones to esters (lactones) upon their treatment with peroxy reagents, was the most obvious choice for the transformation of ring B. We decided to apply the simplest oxidant, that is, m-chloroperbenzoic acid (MCPBA) to introduce an oxygen atom into the cyclohexane ring B. To avoid possible epoxidation of the double bond in 12, the olefin was converted into cis-diol 13 via simple osmylation, which provided diol 13 as the only product in 72% isolated yield. Its structure was proven by NMR experiments performed on diacetate 13a, in which the NOESY correlation peak between H-4 and H-6 was observed[§] (see [Scheme 1\)](#page-2-0).

Treatment of keto-diol 13 with MCPBA resulted in formation of the Baeyer–Villiger product $[m]z$: 641.2743 = $C_{36}H_{42}O_9Na$ (M+Na⁺)]. From two possible structures 14 and 15 the latter was excluded on the basis of the NMR data. In the COSY spectrum of the Baeyer–Villiger product, the lowest resonance ($\delta = 5.31$ ppm) corresponded to the low field signal at $\delta = 4.05$ ppm.; moreover it was coupled to only one proton (H-2). These results pointed unequivocally at the structure 14 and excluded 15 in which the lowest resonance should be coupled to two high field resonating protons: H-9 and H-5. Although we were unable to assign the configuration at the new stereogenic centre $(C-2)$, structure 14 was proposed (and not the epimer at the C-2 position), which is based on the wellknown fact that the configuration is retained during the Baeyer–Villiger oxidation.^{[14](#page-5-0)}

[†]The synthesis of enantiomerically pure derivatives of this type of sugars was first reported by Herczegh, who used a 'classical' approach, see Ref.

[⁷](#page-5-0). This obvious alternative: cleavage of the C6–C7 bond will not be discussed in this Letter.

 $§$ These protons are in *cis*-relationship and are relatively close to each other. In the alternative structure, no such correlation is possible. Moreover, the stereochemical outcome of this reaction (attack of OsO4 from the 'upper' side) is analogous to one observed earlier by us for a similar bicyclic compound (see Ref. [8](#page-5-0)).

For reasons of clarity, we keep the numbering as in precursors 12 and 13 and the extra oxygen atom in ring \bf{B} in compound 14 is assigned the ' $1a$ ' label.

Scheme 1. Reagents and conditions: (i) THF, H_2O , H_3O , NMO, OsO_4 (cat.), 72%; (ii) Ac₂O, py, quant.; (iii) MCPBA, NaHCO₃, CH₂Cl₂, 65%.

2.2. Approach to highly oxygenated decalis: functionalization of the allylic C6–C8 fragment in 12

As pointed out in the Introduction (see also Refs. [1 and](#page-5-0) [10\)](#page-5-0), the most obvious choice for the oxidation of the allylic methylene group, that is, direct oxidation or base-catalyzed rearrangement of the corresponding epoxides, is not applicable to our bicyclic compounds. A solution to this problem is provided by the highly regioselective opening of the oxirane ring in the epoxides derived from the bicyclic olefin 16. Reaction of the nucleophile (ACO^-, N_3^-) with epoxides 17 and 20 furnished the corresponding trans-diols or azidoalcohols 18 and 19 in high yields (Fig. 4).[8](#page-5-0)

Figure 4. Model study on the oxidation of the precursors of highly oxygenated decalins.

Recently we have applied the well established selenium methodology^{[15](#page-5-0)} for the opening of the epoxide ring in 17, which provided the expected allylic alcohol 21 in good yield (Scheme 2).^{[16](#page-5-0)}

The isomeric (to 17) epoxide 20 might be a convenient precursor of olefin 22, a useful building block for the stereoselective synthesis of carbobicyclic derivatives bearing a quaternary carbinol centre. Thus, treatment of oxirane 20 with the phenylselenide anion followed by oxidative work-up provided alcohol 22 in good yield (Scheme 2). Its structure was thoroughly verified by NMR analysis performed on its acetate derivative 22a. The signal of the H-7 proton occurring at $\delta = 5.30$ ppm correlated to the olefinic H-6 (δ = 6.06 ppm) and two high field resonances (δ = 1.85 and 1.45 ppm) in the COSY spectrum of 22a. These two high field protons correlated to the secondary carbon at $\delta = 29.2$ ppm proving the presence of the CH₂– group at the C-8 position.

Compound 22 is (or should be) a convenient precursor of highly oxygenated decalins bearing a quaternary carbinol centre (at C-5) or carbasugars (via a cleavage of the double bond followed by reduction of the resulting ketone into the 5 -CH₂ group).

Exploring the synthetic potential of the 'allyltin approach' to highly oxygenated carbobicycles, we turned our attention to allylic alcohol 21 with the (R) -configuration at the carbinol (C-6) centre. Besides the obvious functionalization of the double bond (osmylation, aminohydroxylation, epoxidation) this compound can be converted into the C-6 epimer by simple inversion of the configuration at this stereogenic centre, which will open a route to new, configurationally different, highly oxidized decalins. Treatment of alcohol 21 with the Mitsunobu reagent^{[17](#page-5-0)} gave quite unexpectedly two products: the desired S_N^2 product 24a and the rearranged S_N2' alcohol 23 ([Scheme 3\)](#page-3-0), although it is known that the S_N2' reaction under the Mitsunobu conditions are very rare.[18,19](#page-5-0)

Scheme 2. Reagents and conditions: (i) (1) PhSe–SePh, NaBH₄, EtOH; (2) $H₂O₂$, 71% overall.

Scheme 3.

The structure of the rearranged product 23 was proven by NMR experiments. In the COSY spectrum, the correlation peak between the olefinic (H-6) and high-field H-5 signals was observed. The relative *cis*-relationship between the H-8 and H-2 was assigned by the corresponding correlation peak in the NOESY spectrum (Fig. 5). This showed that

Figure 5. Assignment of the configuration of the rearranged Mitsunobu product.

Figure 6. The example of the allylic rearrangement under Mitsunobu conditions.

the attack of the carboxylic acid $(ArCOOH = p\text{-nitrobenz-}$ oic acid) on the allylic system (as activated by the Mitsunobu reagent alcohol 21) occurred on the opposite site (inversion) in an S_N2 mode and from the same side (retention) in an S_N2' mode (Scheme 3).

The addition of the nucleophile from the same side as the pre-existing OH grouping is in agreement with the recent observation on such allylic Mitsunobu rearrangements, observed during the synthesis of conduritols (Fig. 6).^{\parallel [,19](#page-5-0)}

The expected S_N2 compound, alcohol 24, was obtained as acetate 24b, however, as the only product in the reaction of the corresponding mesylate with acetate anion. Compound 24b was different from acetate 21-Ac; in the NOESY spectrum of 24b no correlation between the H-6 and H-4 was seen (which were visible in the spectrum of the starting carbinol 21).

Work on functionalization of all allylic alcohols 21 and 22 and those obtained after hydrolysis of esters (23 and 24) is currently in progress and will be reported in due course.

3. Conclusion

We have explored the route leading to highly functionalized decalins and cyclohexanes from sugar allyltin derivatives. We have observed that the Mitsunobu reaction, proceeding in most cases in an S_N^2 mode, afforded a rearranged $(S_N 2')$ product when applied to the bicyclic allylic alcohols. Such rearrangements, although reported in the literature, are rare under the Mitsunobu conditions.

4. Experimental

4.1. General

NMR spectra were recorded with a Bruker AM 500 spectrometer for solutions in CDCl₃ (internal Me₄Si). Most of the resonances were assigned by COSY $(^1H-^1H)$ and/ or HETCOR and DEPT correlations. The ¹H- and ¹³Caromatic resonances occurring at the typical δ values were omitted for simplicity. Mass spectra were recorded with an ESI/MS Mariner (PerSeptive Biosystem) mass spectrometer. Optical rotations were measured with a Digital Jasco polarimeter DIP-360 ($\lambda = 589$ nm) for solutions in CHCl₃ ($c = 1$) at room temperature. Column chromatography was performed on silica gel (Merck, 70-230 or 230–400 mesh). Methylene chloride was distilled from $CaH₂$ and THF from potassium prior to use. Organic solutions were dried over anhydrous magnesium or sodium sulfate.

This rearrangement is, however, rather an exception. For similar cyclic allylic alcohols the Mitsunobu inversion proceeded without any rearrangement (see Ref. [20\)](#page-5-0).

4.2. (2R,3S,4R,5R,6S,7R,9S,10R)-{2,3,4-Tri-O-benzyl-6,7 dihydroxy-1-keto-9-[(1′*R*)-5,5-dimethyl-2,4-dioxolane-1′yl]}bicyclo[4.4.0]decane 13

Osmylation of the double bond was conducted under stan-dard catalytic conditions.^{[21](#page-5-0)} To a solution of olefin 12^{11} 12^{11} 12^{11} $(350 \text{ mg}, \overline{0.62 \text{ mmol}})$ in THF $(10 \text{ mL}),$ tert-butanol (1 mL) and water (0.1 mL), N-methyl morpholine N-oxide (100 mg) and osmium tetraoxide (3.1 mL of ca 2% solution in toluene) were added. The mixture was stirred for 24 h at room temperature, and partitioned between brine (15 mL) and ethyl acetate (20 mL). The organic layer was separated and the aqueous one extracted with ethyl acetate $(2 \times 20 \text{ mL})$, and the combined organic solutions were dried and concentrated. Purification of the crude product by column chromatography (hexane–ethyl acetate, 2:1) afforded the title diol 13 as an oil (267 mg, 72%). This compound was further characterized as a diacetate: $\bar{a}_{D} = +17.7$; HRMS [ESI] m/z : 709.2990 [C₃₆H₄₄O₈Na $(M+Na^{+})$ requires 709.2983]. Anal. Calcd for $C_{40}H_{46}O_{10}$: C, 69.95; H, 6.75. Found: C, 69.62, H, 6.90%. ¹H NMR δ : 5.71 (\sim t, $J_{5,6} = J_{6,7}$ 2.7, H-6), 4.98 (m, H-7), 4.62 (d, $J_{2,3}$ 10.0, H-2), 4.00 (m, H-2'a, H-4), 3.89 (m, H-1'), 3.66 (dd, $J_{3,4}$ 8.8, H-3), 3.55 (m, H-2'b), 2.70 (dd, $J_{9,10} = J_{5,10}$ 5.0, H-10), 2.28 (m, H-9), 2.10 (m, H-5), 2.08 and 1.99 $[2 \times s, 2 \times C(O)CH_3]$, 1.58 (m, H-8); ¹³C NMR δ : 205.4 (C-1), 169.9 and 169.4 $[2 \times OC(O)CH_3]$, 109.6 (C-3'), 86.7 (C-3), 84.9 (C-2), 78.6 (C-1'), 77.1 (C-4), 76.1, 75.6 and 72.8 $(3 \times OCH_2Ph)$, 68.0 (C-7), 67.6 (C-2'), 66.8 (C-6), 51.3 (C-10), 43.6 (C-5), 40.0 (C-9), 27.0 (C-8), 26.7 and 25.9 $[C(CH_3)_2]$, 20.9 and 20.8 $[2 \times C(O)CH_3]$.

4.3. (2S,3S,4R,5R,6S,7R,9S,10R)-{2,3,4-Tri-O-benzyl-6,7 dihydroxy-1-keto-9-[(1′*R*)-5,5-dimethyl-2,4-dioxolane-1′yl]}bicyclo[4.5.0]-2-oxa-undecane 14

Diol 13 (75 mg 0.12 mmol) was dissolved in CH_2Cl_2 (10 mL) to which NaHCO₃ (16 mg) and *m*-chloroperbenzoic acid (89 mg, 55% purity) were added and the mixture stirred for 16 days at room temperature. The organic layer was washed with saturated $NAHCO₃$ solution, water, dried, concentrated and the product was isolated by column chromatography (hexane–ethyl acetate, 4:1) to yield lactone 14 (48 mg, 65%) as an oil; [α] $_D = -5.1$; HRMS *m*/z: 641.2743 $[C_{36}H_{42}O_9Na (M+Na^{+})$ requires: 641.2721]. ¹H NMR δ : 5.31 (d, $J_{3,4}$ 3.8, H-2), 4.52 (m, H-1'), 4.14 (m, H-2'a), 4.05 (dd, J4,5 3.8, H-3), 3.88 (m, H-4, H-7), 3.64 (m, H-6, H-10, H-2'b), 2.44 (m, H-5), 2.30 (m, H-8a), 2.05 (m, H-9), 1.61 (m, H-8b), 1.30 and 1.26 $[\text{C}(CH_3)_2]$; ¹³C NMR δ : 172.6 (C-1), 109.1 (C-3'), 104.6 (C-2), 84.4 (C-3), 80.2 (C-4), 75.9 (C-1'), 75.2, 74.3, 71.3 ($3 \times OCH_2Ph$), 73.8 (C-6), 68.5 (C-2'), 38.8 (C-10), 37.8 (C-9), 37.0 (C-5), 30.0 (C-8), 27.7 and 27.1 $[**C**(**CH**₃)₂].$

4.4. (1R,2R,3S,4R,7S,9S,10R)-{1,2,3,4-Tetra-O-benzyl-7 hydroxy-9-[(1'*R*)-5,5-dimethyl-2,4-dioxolane-1'-yl]}bicyclo-[4.4.0]dec-5, 6-ene 22

To a solution of diphenyldiselenide (58 mg, 0.19 mmol) in anhydrous ethanol (5 mL), sodium borohydride was added in portions (31 mg, 0.82 mmol) and the mixture was stirred for 15 min until the yellow colour disappeared. Then a solution of epoxide 20 (177 mg, 0.26 mmol) in THF (2 mL) was added and the mixture was boiled at reflux for 2 h. After cooling to 5 $\mathrm{^{\circ}C}$ (ice bath), hydrogen peroxide (30% in water, 1 mL) was added, and the mixture was stirred for 24 h at room temperature, and quenched with saturated sodium carbonate (5 mL). Water (100 mL) was added, and the product was extracted with diethyl ether $(3 \times 15 \text{ mL})$. The organic fraction was washed with brine, dried, concentrated and the product was isolated by column chromatography (hexane–ethyl acetate, 2:1) to give olefin **22**(126 mg; 71%) as an oil. MS m/z : 699.2 $[C_{43}H_{48}O_7 \ (M+Na^+)]$. Anal. Calcd for $C_{43}H_{48}O_7$: C, 76.31; H, 7.15. Found: C, 76.20; H 7.24. This compound was further characterized as acetate 22a: $[\alpha]_D = +19.9$; ¹H NMR δ: 6.07 (m, H-6), 5.29 (m, H-7), 4.25, 4.11, 3.80 $(4H, H-1, H-2, H-3, H-4), 3.94$ (m, H-2'), 3.89 (m, H-1'), 3.50 (t, $J_{1'2'} = J_{2'a,2'b}$ 8.1, H-2'), 2.50 (m, H-10), 2.21 (m, H-9), 2.00 [s, 3H, C(O)CH3], 1.84 (m, H-8a), 1.45 (m, H-8b); ¹³C NMR δ : 170.8 [OC(O)CH₃], 140.7 (C-5), 123.2 $(C-6)$, 108.9 $[C(CH₃)₂]$, 81.7, 79.5, 78.6 and 77.1 $(C-1, C-1)$ 2, C-3, C-4), 78.5 (C-1'), 67.7 (C-2'), 66.0 (C-7), 38.8 (C-10), 33.5 (C-9), 29.2 (C-8), 25.8 and 25.7 [C(CH3)2], 21.3 $[CO)CH₃$].

4.5. Mitsunobu reaction of alcohol 21

A solution of 21 (64 mg, 0.09 mmol), triphenylphosphine (52.5 mg, 0.2 mmol), diisopropylazadicarboxylate (40.5 mg, 0.2 mmol) and 4-nitrobenzoic acid (33.4 mg, 0.2 mmol) in anhydrous THF (10 mL) was stirred for 3 days at room temperature, and then boiled at reflux for 5 h. The reaction mixture was partitioned between saturated sodium bicarbonate (5 mL) and ethyl acetate (10 mL). The organic phase was separated and the aqueous one extracted with EtOAc $(3 \times 10 \text{ mL})$. The combined organic fractions were washed with brine $(2 \times 10 \text{ mL})$, dried, concentrated and the products were isolated by column chromatography (hexane–ethyl acetate, 3:1) to afford 23 as an oil (33 mg; 42%) and 24 (23 mg; 29%).

4.6. (1R,2R,3S,4R,5R,8S,9S,10R)-{1,2,3,4-Tetra-O-benzyl- $8-(4-O-p-nitrobenzoyl)-9-[(1'R)-5,5-dimethyl-2,4-dioxolane-$ 10 -yl]}bicyclo[4.4.0]dec-6,7-ene 23

 $[\alpha]_D = -1.0; \quad \text{HRMS} \quad m/z; \quad \text{848.3381} \quad [\text{C}_{50}\text{H}_{41}\text{NO}_{10}\text{Na}]$ $(M+Na^{+})$ requires 848.3405]. ¹H NMR δ : 6.24 (dd, $J_{5,6}$) 5.5, $J_{6,7}$ 9.9, H-6), 5.86 (dd, $J_{7,8}$ 3.9, H-7), 5.59 (m, H-8), 4.78 (m, H-1'), 3.98 (t, $J_{1,2} = J_{2,3}$ 9.2, H-2), 3.88 (t, $J_{1'2'} = J_{2'a,2'b}$ 8.8, H-2'a), 3.83 (t, J 9.2, H-4), 3.67 (m, H-2'b, H-3), 3.60 (dd, $J_{1,10}$ 4.4, H-1), 2.78 (m, H-9), 2.36 $(m, H-5)$, 2.12 $(m, H-10)$, 1.18 $(m, 6H, [CH_3)_2C]$; ¹³C NMR δ: 164.3 [Ar-C(O)O-], 134.9 (C-6), 125.7 (C-7), 108.6 $[(CH_3)_2C]$, 86.6 (C-3), 83.4 (C-4), 81.96 (C-2), 81.90 (C-1), 75.48 (C-1'), 70.4 (C-8), 65.7 (C-2'), 39.5 (C-9), 38.8 (C-5), 38.0 (C-10), 26.0 and 24.2 $[(CH₃)₂Cl]$.

4.7. (1R,2R,3S,4R,5R,6S,9S,10R)-{1,2,3,4-Tetra-O-benzyl- $6-(4-O-p-nitrobenzoyl)-9-[(1/R)-5,5-dimethyl-2,4-dioxolane-$ 10 -yl]}bicyclo[4.4.0]dec-7,8-ene 24a

 $[\alpha]_D = -4.5;$ HRMS m/z : 848.3442 $[C_{50}H_{41}NO_{10}Na$ $(M+Na^{+})$ requires 848.3405]. ¹H NMR δ : 6.14 (dd, $J_{8,9}$ 2.3, $J_{7,8}$ 10.1, H-8), 5.88 (ddd, $J_{7,9}$ 1.0, $J_{6,7}$ 5.5, H-7), 5.65 (dd, $J_{5,6}$ 1.3, H-6), 5.08 (m, H-1^{*i*}), 3.86 (m, H-2), 3.78 (t, $J_{1',2'a} = J_{2'a,2'b} = 7.2$, H-2'a), 3.59 (m, H-1, H-3), 3.52 (t, H-2'b), 3.42 (m, H-4), 2.80 (m, H-9), 2.00 (m, H-5, H-10), 1.40 and 1.25 (6H, $[\text{CH}_3)_2\text{C}$); ¹³C NMR δ : 163.6 [CH₃C(O)], 132.6 (C-8), 124.0 (C-7), 108.6 [(CH₃)₂C], 86.3 and 83.3 (C-1, C-3), 82.5 (C-2), 77.8 (C-4), 75.5 (C-1'), 68.1 (C-6), 64.6 (C-2'), 41.4 and 33.1 (C-5, C-10), 37.1 (C-9), 32.6 (C-10), 25.9 and 24.5 $[(CH₃)₂Cl]$.

4.8. Synthesis of the 'inverted' alcohol 24 by S_N2 reaction of the corresponding mesylate

To a vigorously stirred solution of alcohol 21 (53.4 mg, 0.079 mmol) in dichloromethane/triethylamine $(3:1 \text{ v/v},$ 4 mL) containing catalytic amounts of DMAP (3 mg), mesyl chloride (90 mg, 0.79 mmol) was added and the mixture was stirred for 4 h. It was then concentrated and the crude mesylate was purified by column chromatography (hexane–diethyl acetate, 3:1) to afford the mesylation product (40 mg). This was dissolved in DMF (5 mL) to which sodium acetate (82 mg, 1 mmol) was added and the reaction mixture was kept at 110° C until disappearance of the starting material (3 days). The mixture was partitioned between water (10 mL) and ether (20 mL), the organic fractions were collected, washed with brine, dried, concentrated and the product was isolated by column chromatography (hexane–ethyl acetate, 3:1) to afford the desired compound 24b (21 mg; 37%) as an oil.

4.9. (1R,2R,3S,4R,5R,6S,9S,10R)-{1,2,3,4-Tetra-O-benzyl- $6-(O\text{-}acyl)\text{-}9-[(1'R)\text{-}5,5\text{-}dimethyl-2,4-dioxolane-1'$ yl]}bicyclo[4.4.0]dec-7,8-ene 24b

 $[\alpha]_{\text{D}} = -13.0;$ m/z : 741.3 $[C_{45}H_{50}O_8Na$ $(M+Na^+)]$. ¹H NMR δ : 6.07 (dd, $J_{8,9}$ 2.8, $J_{7,8}$ 10.2, H-8), 5.86 (ddd, $J_{7,9}$ 1.7, $J_{6,7}$ 5.4, H-7), 5.39 (dd, $J_{5,6} = 2.2$, H-6), 5.03 (m, H-1'), 3.85-3.78 (m, H-2, H-2'), 3.60-3.52 (m, H-4, H-3, H-2'), 3.37 (dd, J 9.1, J 11.5, H-1), 2.75 (m, H-9), 2.06 (m, H-10), 2.01 [s, $CH_3C(O)O$], 1.85 (m, H-5), 1.41 and 1.21 [6H, $(CH_3)_2C$]; ¹³C NMR δ : 169.9 [CH₃C(O)], 131.4 (C-8), 124.7 (C-7), 108.6 [$(CH_3)_2C$], 86.3, 83.7 (C-4, C-3), 82.6 (C-2), 78.1 (C-1), 75.6 (C-1'), 66.3 (C-6), 64.7 (C-2'), 41.4 (C-5), 36.9 (C-9), 32.6 (C-10), 26.0 and 24.5 $[(CH₃)₂C]$, 21.1 $[CH₃C(O)]$.

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References

- 1. Microreview: Jarosz, S.; Gaweł, A. l. Eur. J. Org. Chem. 2005, 3415.
- 2. Selected recent reviews and papers: Arjona, A.; Gomez, A. M.; Lopez, J. C.; Plumet, J. Chem. Rev. 2007, 107, 1919; Long, M.; Ziegler, Th. Eur. J. Org. Chem. 2007, 768; Zhou, J.; Wang, G.; Zhang, L.-H.; Ye, X.-S. Curr. Org. Chem. 2006, 10, 625; Podeschwa, M. A. L.; Plettenburg, O.; Altenbach, H.-J. Eur. J. Org. Chem. 2005, 3101, and 3116; Freeman, S.; Hudlicky, T. Bioorg. Med. Chem. Lett. 2004, 14, 1209, and references cited therein.
- 3. Mehta, G.; Ramesh, S. S. Can. J. Chem. 2005, 83, 581; Mehta, G.; Ramesh, S. S. Tetrahedron Lett. 2001, 42, 1987; Mehta, G.; Ramesh, S. S. Chem. Commun. 2000, 2429.
- 4. Roush, W. R.; Koyama, K.; Curtin, M. L.; Moriarty, K. J. J. Am. Chem. Soc. 1996, 118, 7502.
- 5. Kozłowska, E.; Jarosz, S. J. Carbohydr. Chem. 1994, 13, 889.
- 6. Jarosz, S.; Szewczyk, K.; Zawisza, A. Tetrahedron: Asymmetry 2003, 14, 1715.
- 7. Herczegh, P.; Zsely, M.; Szilagy, L.; Bajza, I.; Kovacs, A.; Batta, G.; Bognar, R. In Cycloaddition Reactions in Carbohydrate Chemistry; Guliano, R. M., Ed.; ACS Symposium Series; OUP: Oxford, 1992; Vol. 494, p 112.
- 8. Jarosz, S.; Skóra, S. Tetrahedron: Asymmetry 2001, 12, 1651.
- 9. Crandall, J. K.; Apparu, M. Org. React. 1983, 29, 345.
- 10. Jarosz, S.; Boryczko, B.; Cmoch, P.; Gomez, A. M.; Lopez, C. Tetrahedron: Asymmetry 2005, 16, 513.
- 11. Jarosz, S.; Skóra, S. Tetrahedron: Asymmetry 2000, 11, 1433.
- 12. Krow, G. R. In Comprehensive Organic Synthesis; Trost, B. M., Fleming, I., Eds.; Pergamon: Oxford, 1991; Vol. 7, p 671.
- 13. Baeyer, A.; Villiger, V. Ber. Dtsch. Chem. Ges. 1899, 32, 3625; Griegee, R. Liebigs Ann. Chem. 1948, 560, 127.
- 14. Turner, R. B. J. Am. Chem. Soc. 1950, 72, 878; Maslow, K.; Brenner, J. J. Am. Chem. Soc. 1953, 75, 2318; See also: Wallace, P.; Warren, S. J. Chem. Soc., Perkin Trans. 1 1992, 3169; Dyker, G.; Grundt, P. Eur. J. Org. Chem. 1999, 1, 323.
- 15. Sharpless, K. B.; Lauer, R. F. J. Am. Chem. Soc. 1973, 95, 2697.
- 16. Jarosz, S.; Nowogródzki, M. J. Carbohydr. Chem. 2006, 25, 139.
- 17. Mitsunobu, O. Synthesis 1981, 1.
- 18. First observation of the S_N^2 Mitsunobu rearrangement see: Grynkiewicz, G.; Burzyńska, H. Tetrahedron 1976, 32, 2109; Other examples up to 1996 see: Jarosz, S.; Kozłowska, E. Carbohydr. Lett. 1996, 2, 213–216, and references cited therein.
- 19. Kwon, Y.-U.; Chung, S.-K. Org. Lett. 2001, 3, 3013.
- 20. Klepper, F.; Jahn, E.-M.; Hickmann, V.; Carell, Th. Angew. Chem., Int. Ed. 2007, 46, 2325; O'Brien, P.; Pilgrim, Ch. D. Org. Biomol. Chem. 2003, 1, 523.
- 21. VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 1973; Jarosz, S. Carbohydr. Res. 1992, 224, 73.